

Research Article



Molecular detection of *Salmonella typhimurium* isolated from canine feces by PCR

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Abstract | This study was conducted to detect five virulence factors include *invA*, *spvC*, *fimA*, *stn* and *Typh* genes on 4 isolates of *S. typhimurium* serotype which isolated previously from 3 puppies and 1 adult dog, this isolates were confirmed by culture, biochemical, Gram stain and serotyped at the National Center of salmonella in the Baghdad / Ministry of Health. The 4 isolates [3 isolated from puppies and 1 from adult] were confirmed by conventional PCR, A five genes were designed for molecular study, these include *invA*, *spvC*, *fimA*, *stn* and *Typh* genes. The results of molecular study showed that all isolates were positive to *Typh* gene, while *invA* gene was detected in two isolates only. One isolate only was having *SpvC* gene. The results of *fimA* gene showed presence of this gene in two isolates, while the *Stn* gene was not detected in in any isolates of *S. typhimurium*. In conclusion, the *Salmonella typhimurium* had been detected in dogs and puppies which can cause an infection in human and PCR system is useful in the detection of virulence of this bacteria.

Keywords | *Salmonella*, Molecular, Canine, *S. typhimurium*, Gene

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INTRODUCTION

Salmonella spp. remains a significant food borne zoonotic pathogen in all regions of the world with *S. typhimurium* as one of the most common serovars causing disease in animals and human (Herrero-Fresno and Olsen, 2018). *Salmonella* is gram-negative, flagellated, non-encapsulated, non-sporulated and aerobic (facultative aerobic) bacilli, they are usually seen as short rods and occasionally develop into a longer pleomorphic form or short coccobacilli when incubated for a long time in the laboratory (Markey et al., 2013).

Salmonella can cause a variety of pathologies, including gastroenteritis, abortions, pneumonia and lethal septicemia in human and animals (Ohl and Miller, 2001). Dogs generally seem to be resistant to *Salmonella* infection and most cases are latent and non-clinical, infected dogs may remain carriers and faecal shedders and thus serve as sources of *Salmonella* for man and other animals (Kozak et al., 2003). A study of Verma et al. (2011) on clinical cases of canine salmonellosis was designed to compare the sensitivity,

specificity and accuracy of PCR to the isolation and characterization methods commonly used in diagnostic laboratories, they indicate that the PCR assay as a simple, rapid, sensitive, reliable and reproducible method for the identification of *Salmonella* that will aid in surveillance, prevention and control of this pathogen.

Makino et al. (1999) established the specific detection to *Salmonella* species by PCR system using *Salmonella* enterotoxin gene (*stn*), This detection was limited to one bacterial cell/ one gram of fecal samples and a minced-meat using enrichment by *Salmonella* enrichment broth or Trypticase soy broth respectively, They concluded that PCR system is useful in the field of the public hygiene.

This study aimed to detect the virulence factors in *Salmonella typhimurium* isolated from canine feces.

MATERIALS AND METHODS

BACTERIAL ISOLATES

Four *S. typhimurium* were isolated in pervious study (Ibra

Table 1: Primers types with their sequences with product size

Primers	Primer sequence (5' to 3')	Product size (bp)	References
invA	F GTG AAA TTA TCG CCA CGT TCG GGC AA	284	Kumar <i>et al.</i> (2008)
	R TCA TCG CAC CGT CAA AGG AAC C		
spvC	F ACT CCT TGC ACA ACC AAA TGC GGA	571	Oliveira <i>et al.</i> (2003)
	R TGT CTT CTG CAT TTC GCC ACC ATC A		
fimA	F CCT TTC TCC ATC GTC CTG AA	85	Naravaneniet <i>al.</i> (2005)
	R TGG TGT TAT CTG CCT GAC CA		
Stn	F CTT TGG TCG TAA AAT AAG GCG	260	Makino <i>et al.</i> (1999)
	R TGC CCA AAG CAG AGA GAT TC		
Typh	F TTG TTC ACT TTT TAC CCC TGA A	401	Olsen <i>et al.</i> , 1995
	R CCC TGA CAG CCG TTA GAT ATT		

Table 2: Number of isolates that carried virulence genes in affected animals

Animals (4)	No. of <i>S. typhimurium</i> isolates(4)	Gene					Total of genes in each isolates
		<i>Typh</i>	<i>fimA</i>	<i>SpvC</i>	<i>InvA</i>	<i>Stn</i>	
1- Puppy with bloody diarrhea	1	☑	☑	☑	☑	-	4
2-Puppy with diarrhea	2	☑	☑			-	2
3-Non diarrheic Puppy	3	☑				-	1
4-Adult dog with bloody diarrhea	4	☑			☑	-	2

him and Yousif, 2018), 3 isolates from puppies with different clinical signs and one isolate from adult dog, these isolates were conducted to cultural, biochemical in the dept. of Internal and Preventive Vet. Medicine and confirmed serologically as *S. typhimurium* in the national center of *Salmonella* in Baghdad.

MOLECULAR STUDY IN ISOLATED BACTERIA BY CONVENTIONAL PCR

Four isolates of *Salmonella typhimurium* were used in the molecular study, these isolates were subjected:

DNA extraction: According to manufacturing procedure. The DNA of *Salmonella* isolates was extracted by using by using (Geneaid DNA Bacterial Kit, USA).

Primers: Five primers in this study were purchased from Bioneer, Koreato detect *salmonella* virulence gene. These primers were prepared according to the information of the company. *invA*, *spvC*, *fimA*, *stn* and *Typh* genes (Table 1).

The concentration and purity of extracting DNA were evaluated by using a Nanodrop (ND-2000, Thermo Fisher scientific, USA). Then gel electrophoresis was used to check the extracted DNA by loading the DNA in 1.5% agarose gel.

PCR mixture component of *invA*, *spvC*, *fimA*, *stn* and

***Typh* genes:** For detecting *invA*, *spvC*, *fimA*, *stn* and *Typh* genes of *Salmonella typhimurium* by PCR, the PCR amplification mixture which used for detection these genes includes master mix (12.5µl), 2 µl of template DNA, 2µl of each forwarded and reversed primers and 6.5µl of nuclease free water to complete the amplification mixture to 25µl.

Thermo- cycler program: The program of thermo-cycle for detection *invA*, *Typh* and *SpvC* genes as follows: One cycle for 3 minutes at 94°C to denature template. It was continued by 35 cycles, each cycle including denaturation for 30 seconds at 94°C, annealing 30 seconds at 63°C, and extension 30 seconds at 72°C. Final extension was done 5 minutes at 72°C.

The program of thermo-cycle for detection of *fimA* gene was done as follows: Initial denaturation ongoing by 1 cycle at 94°C for 3 min. Followed by 35 cycles, each cycle including denaturation, annealing and extension at 94°C for 30 seconds, 56°C for 30 sec., 72°C for 30 sec. The 35 cycles Followed by a final extension for 5 min. at 72°C.

The amplification of *stn* gene was carried out employing same conditions as *invA*, *Typh* and *spvC* genes except annealing at 55°C.

PCR PRODUCT ANALYSIS

This step was used for analyzing the PCR product by using

2% agarose, stained with 0.5 µg/mL ethidium bromide, The PCR products (bands) were visualized by using a UV transilluminator and photographed by using digital camera.

RESULTS

DNA GENOMIC EXTRACTION OF *SALMONELLA*

The four isolates of *Salmonella* were showed compact bands as an indication of DNA extraction successfully in gel electrophoresis (Figure 1).

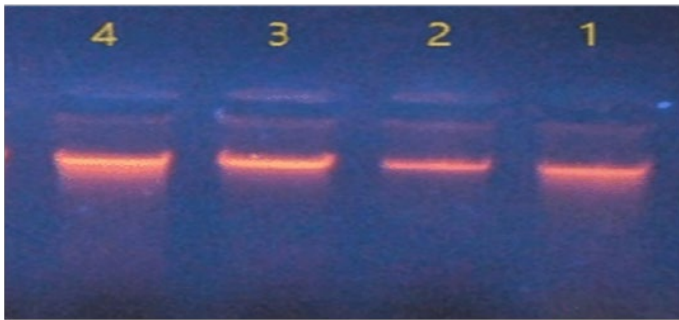


Figure 1: DNA extraction of *Salmonella* isolates.

In Nano drop spectrophotometer the concentration of DNA ranged between 47 – 190 ng/µl and the purity were between 1.6 – 2.1 at absorbance 260/280 nm.

CONFIRMATORY DETECTION OF VIRULENCE GENES BY USING CONVENTIONAL PCR

The results of PCR amplification of the extracted DNA from isolates of *S. typhimurium* on different types of genes used in comparison with DNA marker (100bp ladder) as follows.

Typh gene was amplified DNA at 401bp fragments , All isolates were positive for this gene (Figure 2). While *invA* gene was detected in two isolates only which revealed 284bp (Figure 3).

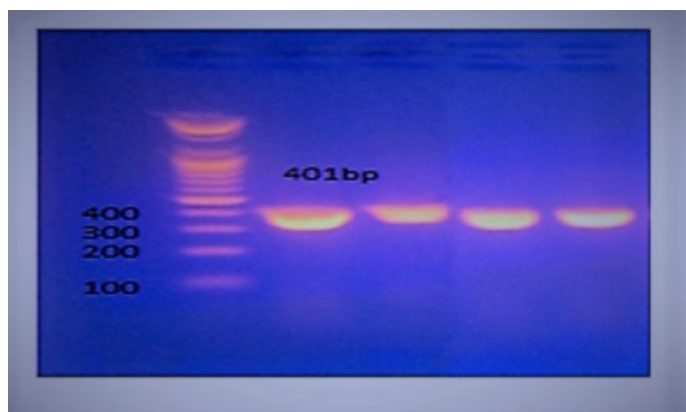


Figure 2: Agarose gel electrophoresis showing amplification of 401 bp fragments of *Typh* gene. Lanes 1,2,3,4 showing positive amplification of *Salmonella typhimurium*

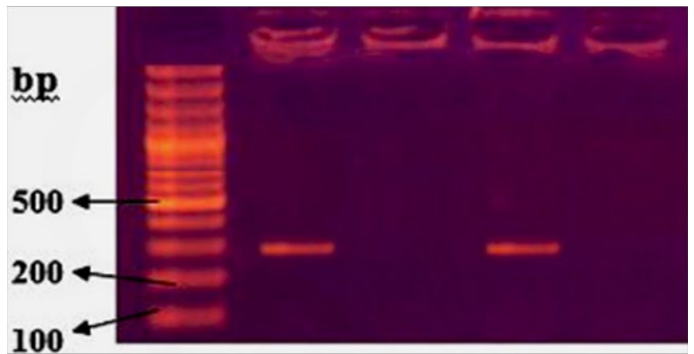


Figure 3: Agarose gel electrophoresis showing amplification of 284bp fragments of *invA* gene. Lanes 1,3 showing positive amplification of *Salmonella typhimurium*

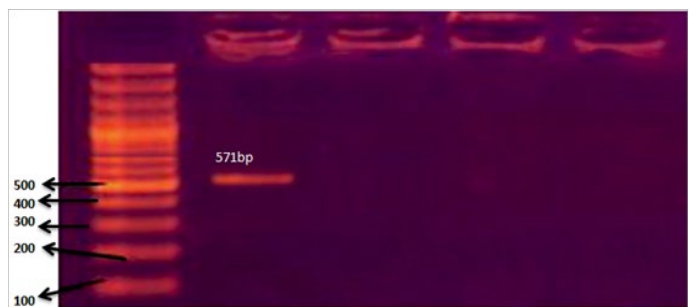


Figure 4: Agarose gel electrophoresis showing amplification of 571bp fragments of *SpvC* gene. Lane 1 showing positive amplification of *Salmonella typhimurium*.

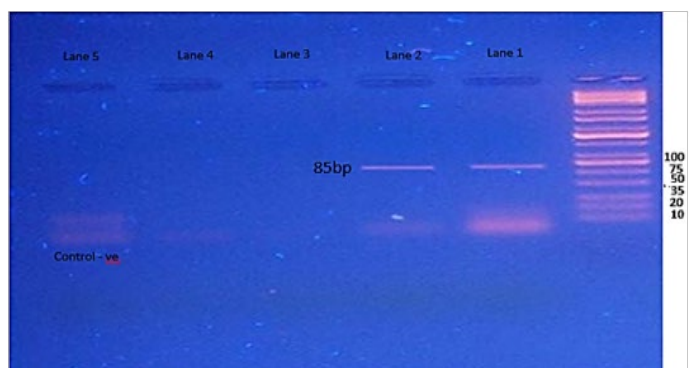


Figure 5: Agarose gel electrophoresis showing amplification of 85bp fragments of *fimA* gene. Lanes 1,2 showing positive amplification of *Salmonella typhimurium*.

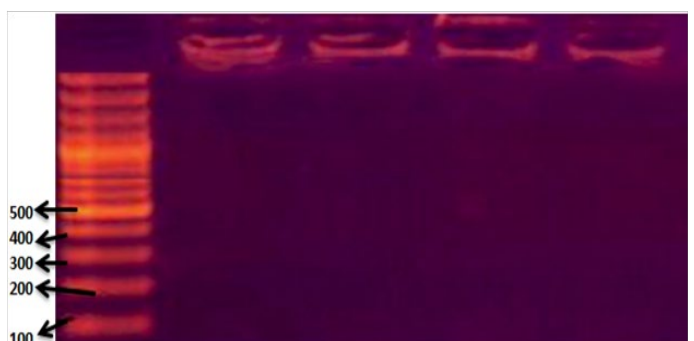


Figure 6: Agarose gel electrophoresis showing amplification of 280bp fragments of *stn* gene. Lanes 1,2,3,4 showing negative amplification of *Salmonella typhimurium*.

The *SpvC* gene showed amplification of 571bp fragments, one isolate only was have this gene (Figure 4). The results of *fimA* gene showed amplification of 85bp fragments in two isolates of *S. typhimurium* (Figure 5), while the *Stn* gene was not detected in in any isolates of *S. typhimurium*. (Figure 6).

DISTRIBUTION OF VIRULENCE GENES AMONG INFECTED ANIMALS

The presented results in a Table 2 revealed the distribution of different genes in each isolate of *Salmonella typhimurium*. The first isolate from Puppy with bloody diarrhea carried 4 genes (*Typh*, *fimA*, *SpvC* & *InvA*), second isolate from -Puppy with diarrhea carried 2 genes (*Typh* & *fimA*), the third isolate from-Non diarrheic Puppy was carried *Typh* gene only. While the *Salmonella* isolate from adult dog was possessed 2 genes (*Typh*, & *InvA*).

DISCUSSION

All the results of culturing, gram stain and biochemical tests are similar to the morphological characters were compatible with the information mentioned by (Markey et al., 2013; Mahon et al., 2015).

In-vitro amplification of DNA by the PCR method is a powerful tool in microbiological diagnostics (Malorny et al., 2003). Several genes have been used to detect *Salmonella* in natural environmental samples as well as food and faecal samples.

The current result showed that the PCR was a sensitive and specific method as compared with bacterial culture and this result was in agreement with a result of Kurowski et al. (2002) who used PCR technology to develop a highly sensitive and specific diagnostic assay for the detection of *Salmonella* spp in fecal dogs specimens, they found that Use of the *spaQ* primer-probe set resulted in a relative sensitivity of 100% and a specificity of 98.2%, compared with bacterial culture results when tested on 299 clinical fecal specimens.

Also, the results were incompatible with result of Parungao et al. (2010) who used conventional isolation procedures and PCR for the presence of *Salmonella* spp. in feces of dog, they detected *salmonella* in both conventional microbiological procedures (1 out of 62 or 1.6%) and PCR (26 out of 62 or 42%) of the samples tested, they concluded that apparently healthy dogs can be carriers and shedders of potentially zoonotic salmonellae, and that PCR can be an effective means of detecting the said pathogen in canine fecal samples.

Virulence chromosomal genes including, *invA*, *invE*,

himA, *phoP* are target genes for PCR amplification of *Salmonella* species (Jamshidi et al., 2009).

The *invA* gene of *Salmonella* contains sequences unique to this genus and has been proven as a suitable PCR target, with potential diagnostic applications (Rahn et al., 1992). Amplification of this gene now has been recognized as an international standard for detection of *Salmonella* genus (Malorny et al., 2003).

The presented results showed a high prevalence of *invA* gene at percent 50%, this result agree with result of Hashemi et al. (2014) who aimed to find out the prevalence rate of *Salmonella* infection in apparently healthy cats and dogs in Iran by using PCR by using *invA* gene, they found that prevalence rates of *Salmonella* were 18 and 22% in cats and dogs respectively, also incompatible with Srisanga et al. (2017) who showed that most canine *Salmonella* isolates harbored *invA* (100%), and agree with Torkan et al. (2015) who reported a high prevalence of *invA* gene in *Salmonella* isolates, this result indicates that pet dogs and cats serve as reservoirs of invasive *Salmonella*.

Also, Chaudhary et al. (2015) isolate *Salmonella* Enteritidis and Typhimurium from pork and slaughter house environment in Ahmedabad, Gujarat out of a total of 270 samples, 37 (13.70%), All *Salmonella* serovars produced *invA* gene, *fimA* and amplicon for enterotoxin (*stn*) gene whereas 30 isolates possessed 310 bps *spvR* gene, but no isolate possessed *spvC* gene.

Kshirsagar et al. (2014) found a high percent *invA*, *stn* and *fimA* genes associated with *Salmonella* isolated from the retail meat market of raw buffalo meat and offals viz. liver, lung, muscle, intestine and ground beef in Gujarat India. In Iraq, Al-Zubaidy et al. (2015) detected the *invA* gene in the *Salmonella* spp. isolated from the cows

A total of 37 *Salmonella* isolates of 11 different serotypes and rough type of human and animal clinical cases and meat samples were studied for the presence of 8 virulence determinants including 4 virulence genes and 4 toxic factors, All *Salmonella* isolates harbored *invA* and *stn* genes (Singh et al., 2013), these results were didn't agree with our result about the *stn* gene which not presented in the current study. Ulaya (2013) showed that the *invF* gene was (16.7%) as well as the *sipC* gene (8%) the *hilD* (5.6%) gene belonging to *Salmonella* in Zambia from dogs.

CONCLUSION

Salmonella isolated from companion animals (dogs and puppies) and these have a many virulence factors which can transmit to human and causing infection.

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CONFLICT OF INTEREST

The author declares that they have no competing interests.

AUTHORS CONTRIBUTION

All the authors contributed equally.

REFERENCES

- Al-Zubaidy AAN, Yousif AA, Al-Graibawi MAA, Darkhan J (2015). Detection of invasion gene *invA* in *Salmonella* spp. isolated from slaughtered cattle by PCR method. *Iraq J. Vet. Med.* 39(1): 128-133.
- Chaudhary JH, Nayak JB, Brahmabhatt MN, Makwana PP (2015). Virulence genes detection of *Salmonella* serovars isolated from pork and slaughterhouse environment in Ahmedabad, Gujarat. *Vet. World.* 8(1): 121. <https://doi.org/10.14202/vetworld.2015.121-124>
- Hang P, Paudyal N, Li X, Fang W, Yue M (2018). Multiple food-animal-borne route in transmission of antibiotic-resistant *Salmonella* Newport to humans. *Front. Microbiol.* 9: 23. <https://doi.org/10.3389/fmicb.2018.00023>
- Hashemi A, Baghbani-Arani F (2015). The effective differentiation of *Salmonella* isolates using four PCR-based typing methods. *J. Appl. Microbiol.* 118(6): 1530-1540. <https://doi.org/10.1111/jam.12805>
- Herrero-Fresno A, Olsen J (2018). *Salmonella Typhimurium* metabolism affects virulence in the host - A mini-review. *Food Microbiol.* 71:98-110. <https://doi.org/10.1016/j.fm.2017.04.016>
- Jamshidi A, Bassami MR, Afshari-Nic S (2009). Identification of *Salmonella* spp. and *Salmonella* Typhimurium by a multiplex PCR-based assay from poultry carcasses in Mashhad-Iran. *Iran J. Vet. Res.* 3: 43-48.
- Kozak M, Horosova K, Lasanda V, Bilek J, Kyselova J (2003). Do dogs and cats present a risk of transmission of salmonellosis to humans?. *Bratislavské lekárske listy.* 104(10): 323-328.
- Kshirsagar DP, Singh S, Brahmabhatt, MN, Nayak JB (2014). Isolation and Molecular Characterization of Virulence-Associated Genes of *Salmonella* from Buffalo Meat Samples in Western Region of India. *Israel J. Vet. Med.* Vol. 69 (4): 228-233.
- Kumar K, Saklaini, AC, Singh S, Singh VP (2008). Evaluation of specificity for *invA* gene PCR for detection of *Salmonella* spp. In Proceeding of V IIth Annual Conference of Indian Association of Veterinary Public Health Specialists (IAVPHS).
- Kurowski PB, Traub-DargatzJL, Morley PS, Gentry-Weeks CR (2002). Detection of *Salmonella* spp in fecal specimens by use of real-time polymerase chain reaction assay. *American J. Vet. Res.* 63(9): 1265-1268. <https://doi.org/10.2460/ajvr.2002.63.1265>

- Ibrahim WS, Yousif AA (2018). Molecular detection of *Salmonella typhimurium* and some virulence genes from canine feces by PCR. *Online J. Vet. Res.* ©.22 (9):739-743.
- Mahon C, Lehman D, Manuselis G (2015). *Textbook of Diagnostic Microbiology.* Fifth edition, Saunders, Elsevier. 425.
- Makino SI, Kurazono H, Chongsangum M, Hayashi H, Cheun HI, Suzuki S, Shirahata T (1999). Establishment of the PCR system specific to *Salmonella* spp. and its application for the inspection of food and fecal samples. *J. Vet. Med. Sci.* 61(11): 1245-1247. <https://doi.org/10.1292/jvms.61.1245>
- Malorny B, Hoorfar J, Bunge C, Helmuth R (2003). Multicenter validation of the analytical accuracy of *Salmonella* PCR: towards an international standard. *Appl. Environ. Microbiol.* 69(1): 290-296. <https://doi.org/10.1128/AEM.69.1.290-296.2003>
- Markey B, Leonard F, Archambault M, Cullinane A, Maguire D (2013). *Clinical veterinary microbiology.* Second edition, Mosby, Elsevier. 246-249.
- Naravaneni R, Jamil K (2005). Rapid detection of food-borne pathogens by using molecular techniques. *J. Med. Microbiol.* 54(1): 51-54. <https://doi.org/10.1099/jmm.0.45687-0>
- Ohl ME, Miller SI (2001). *Salmonella: a model for bacterial pathogenesis.* *Annual Rev. Med.* 52(1): 259-274. <https://doi.org/10.1146/annurev.med.52.1.259>
- Oliveira SDD, Rodenbusch CR, Michael GB, Cardoso MI, Canal CW, Brandelli A (2003). Detection of virulence genes in *Salmonella* Enteritidis isolated from different sources. *Brazilian J. Microbiol.* 34: 123-124. <https://doi.org/10.1590/S1517-83822003000500042>
- Olsen JE, Aabo S, Rasmussen OF, Rossen L (1995). Oligonucleotide probes specific for the genus *Salmonella* and for *Salmonella typhimurium*. *Letters Appl. Microbiol.* 20(3): 160-163. <https://doi.org/10.1111/j.1472-765X.1995.tb00416.x>
- Parungao SP, Gordoncillo MJ N, Baldrias LR, Ramirez TJ (2010). Isolation and molecular detection of *Salmonella* spp. from the feces of apparently healthy dogs. *Philippine Journal of Veterinary Medicine,* 47(2): 73-77.
- Rahn K, De Grandis SA, Clarke RC, McEwen SA, Galan JE, Ginocchio C, Gyles CL (1992). Amplification of an *invA* gene sequence of *Salmonella typhimurium* by polymerase chain reaction as a specific method of detection of *Salmonella*. *Mole. Cellul. Probes.* 6(4): 271-279. [https://doi.org/10.1016/0890-8508\(92\)90002-F](https://doi.org/10.1016/0890-8508(92)90002-F)
- Singh R, Yadav AS, Tripathi V, Singh RP (2013). Antimicrobial resistance profile of *Salmonella* present in poultry and poultry environment in north India. *Food Control.* 33(2): 545-548. <https://doi.org/10.1016/j.foodcont.2013.03.041>
- Srisanga S, Angkititraku S, Sringam P, Le Ho PT, Vo AT, Chuanchuen R (2017). Phenotypic and genotypic antimicrobial resistance and virulence genes of *Salmonella enterica* isolated from pet dogs and cats. *J. Vet. Sci.* 18(3): 273-281. <https://doi.org/10.4142/jvs.2017.18.3.273>
- Torkan S, Khamesipour F, Anyanwu MU (2015). Detection of virulence and antibacterial resistance genes in *Salmonella* isolates from diarrhoeic dogs in Iran. *Rev. Med. Vet.* 166: 221-228.
- Ulaya W (2013). Determination of virulence factors in *Salmonella* isolates of human, poultry and dog origin in Lusaka district, Zambia. A thesis submitted to the University of Zambia in fulfillment of the requirements of the degree of Master of Science in Microbiology.

- Verma AK, Sinha DK, Singh BR (2011). Detection of Salmonella from clinical samples of dogs by PCR. Indian J.

Anim. Sci. 81(6): 552.