Research Article



Scanning Electron Microscopic Study of the Anthelmintic Effects of Some Anthelmintic Drugs on Poultry Nematode, *Ascaridia galli*

Pawi Bawitlung Lalthanpuii, Kholhring Lalchhandama*

Department of Life Sciences, Pachhunga University College, Aizawl 796001, Mizoram, India.

Abstract | Praziquantel/pyrantel pamoate (PZQ/PTP) combination and albendazole (ABZ) are two commercially available anthelmintic drugs most commonly used for the treatment of different helminth infections. Their anthelmintic activity was studied *in vitro* against the poultry nematode, *Ascaridia galli*. Although both the drugs were effective, ABZ showed most prominent activity. Scanning electron microscopy of the nematodes revealed structural damaged induced by the drugs. Shrinkage of the cuticle, collapse of the lips, and formation of cuticular folds were the defining anthelmintic effects. Comparison based the drug efficacy and anthelmintic effects indicates that ABZ can be the drug of choice for helminthiasis due to nematodes.

Keywords | Anthelmintic drug, Cuticle, Helminthiasis, Nematode, Scanning electron microscopy

Received | April 13, 2020; Accepted | June 21, 2020; Published | July 07, 2020

*Correspondence | Kholhring Lalchhandama, Department of Life Sciences, Pachhunga University College, Aizawl 796001, Mizoram, India; Email: chhandama@pucollege.edu.in

Citation | Lalthanpuii PB, Lalchhandama K (2020). Scanning electron microscopic study of the anthelmintic effects of some anthelmintic drugs on poultry nematode, *Ascaridia galli*. Adv. Anim. Vet. Sci. 8(8): 788-793.

DOI | http://dx.doi.org/10.17582/journal.aavs/2020/8.8.788.793

ISSN (Online) | 2307-8316; ISSN (Print) | 2309-3331

Copyright © 2020 Lalthanpuii et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

Telminthiasis turns out to be the champion of all Linfectious diseases. It remains the primary factor of morbidity and mortality not only in humans but also in livestock animals. Schistosomiasis, which accounts for 290 million cases in humans as of the latest World Health Organization report (WHO, 2020a), surpasses malaria at 228 million cases as the most prevalent parasitic infection. Nematode infection is estimated to be more prevalent and soars at 1.5 billion cases, i.e. infecting almost a quarter of human population (WHO, 2020b). These nematodes are the major cause of morbidity and morbidity-associated mortality (Kyne et al., 2019). The situation is worsened by rapid and widespread evolution of drug resistance in all major helminths of veterinary importance (Schulz et al., 2018). With reports of total failure of deworming drugs, livestock industry is facing massive economic challenges (Tinkler, 2019).

Albendazole (ABZ) and praziquantel (PZQ) are the most popular anthelmintic drugs for their high efficacy

and broad-spectrum activity. They remain the drugs of choice for over three decades for helminthiasis in humans and livestock animals (Hong, 2018). They are also at the frontline medication in debilitating helminth infections such as neurocysticercosis (Garcia et al., 2016), and echinococcosis (Siles-Lucas et al., 2018). However, PZQ is effective against cestodes and trematodes, and not on nematode. In contrast, pyrantel pamoate (PTP) is a nematode-specific drug commonly used in veterinary prescriptions and normally in combination with other drugs. In so far as efficacy is concerned, albendazole alone is effective against all important helminths and produces the highest cure rate (Moser et al., 2017, 2019).

Ascaridia galli Schrank, 1788, is by far the most prevalent and thereby the most important parasite of poultry and wild birds. It is an intestinal nematode causing a range of health problems and pathological complications such as anaemia, droopiness, emaciation and diarrhoea (Brar et al., 2016). The primary effect is loss of nutrient absorption, and severe condition that can lead to death (Gauly et al., 2005). In hens, infection causes loss of appetite, weight loss,



reduced egg production and death (Kilpinen, 2005). It also influences increased chances of bacterial infections, which usually add to the gross pathogenicity (Ruhnke et al., 2017; Foka et al., 2019). *A. galli* are also the only parasites known to infect eggs of their hosts (Bharat et al., 2017). There is no standard anthelmintic drug for the treatment of poultry helminthiasis. It is therefore interesting to study the effects of ABZ and PZQ on this nematode.

MATERIALS AND METHODS

CHEMICALS AND DRUGS

All reagents used were of standard analytic grades manufacture by HiMedia Laboratories Pvt. Ltd., Mumbai, India. Tetramethylsilane, osmium tetroxide and sodium cacodylate were procured from Merck India, Mumbai, India. Kiwof® (20 mg praziquantel+230 mg pyrantel pamoate) was a product of Sava Medica Limited, Pune, Maharashtra, India. Zenlee® (400 mg albendazole) was a product of UNI-PEX Pharmaceutical Private Limited, New Delhi, India.

IN VITRO SURVIVAL TEST

Live Ascaridia galli Schrank, 1788, were collected from the intestines of freshly slaughtered local poultry, Gallus gallus Linnaeus, 1758. The study approved by the Institutional Ethics Committee of Pachhunga University College (PUC-IAEC-2016-Z2 of 10/08/2016). Anthelmintics were prepared in increasing concentrations of 0.5, 1, 2, 5, 10 and 20 mg/ml in 0.9% neutral phosphate-buffered saline (PBS) supplemented with 1% dimethylsulfoxide (DMSO). The highest dose corresponded to that of albendazole (ABZ) at 10 mg/kg body weight of poultry. Kiwof° was adjusted to 20 mg/ml of pyrantel pamoate (PTP), as praziquantel (PZQ) is not an antinematodal drug. In vitro condition was maintained at 37±1°C in a glass-chambered biological incubator.

Negative control consisted of nematodes maintained in only PBS with DMSO. A set of two worms were tested for each dose, and each test was done in triplicates. Death was recorded when the worms failed to show movement even under agitation in lukewarm PBS (45°C). Data were recorded as statistical means \pm standard deviation. Drug efficacy was normalised against the survival time of the control. Significance of the activity was calculated using Student's *t*-test. One-way ANOVA was used to compare the efficacy of the two drugs. The level of significance was considered at ρ < 0.05.

SCANNING ELECTRON MICROSCOPY

Nematodes treated the highest dose, i.e. 20 mg/ml of the anthelmintic drugs were processed for scanning electron microscopy. They were fixed in 4% neutral-buffered

formaldehyde at 4°C for 4 hours. Secondary fixation was done in 1% osmium tetroxide at 4°C for 1 hour. The specimens were dehydrated in acetone. They were then treated with tetramethyl silane for 15 minutes. They were then dried in an air-drying chamber, which was maintained at 25°C. The dried specimens were fixed on metal stubs, and then sputter coated with gold in an ion-sputtering chamber (JFC 1100, JEOL Ltd., Tokyo, Japan). Finally, they were examined under a scanning electron microscope (JSM-6360, JEOL Ltd., Tokyo, Japan) with an electron accelerating voltage of 20 kV.

RESULTS

Both PZQ/PTP and ABZ were effective against *A. galli* and showed dose-dependent efficacy at the concentrations of 0.5, 1, 2, 5, 10 and 20 mg/ml as shown in Table 1. PZQ/PTP took 96.73 \pm 1.56, 90.44 \pm 1.58, 86.98 1.09, 76.01 \pm 1.28, and 62.28 \pm 1.11 h, to kill all the nematodes at the different concentrations, while ABZ took 56.27 \pm 1.25, 37.90 \pm 0.89, 25.17 \pm 0.78, 7.96 \pm 1.21, 1.77 \pm 0.44 h respectively with respect to untreated control. Nematodes in control survived for 226.20 \pm 2.73 h. ABZ was significantly (p < 0.05) more effective than PZQ/PTP at all concentrations tested.

For scanning electron microscopy, untreated (control) A. galli and those treated with 20 mg/ml of the drugs were selected. The untreated nematode exhibits a smooth cylindrical body. Figure 1 shows an anterior (cephalic) region of the body showing three lips that surround the mouth. Eye-like projections on the lips are the sensory organs called papillae. The main body indicated that the body surface, called cuticle, is a marked with a series of transverse line or annulations (Figure 2).



Figure 1: Scanning electron microscopy of the anterior part of an untreated (normal) *A. galli*. Three lips are seen at the bottom. The central hollow area is the mouth. On two lips are small eye-like papillae.

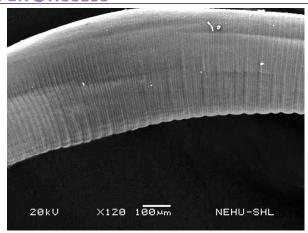


Figure 2: Scanning electron microscopy of the main body of an untreated (normal) *A. galli*. The cylindrical body consists of a series of transverse lines called annulations.

A. galli treated with PZQ/PTP showed severe shrinkage of the body and collapse of the cuticle. In Figure 3, it is seen that the cuticular disintegration caused deflation of the lips. Shrinkage resulted in formation of irregular folds of the cuticle throughout the body (Figure 4). The posterior tail end is also deflated (Figure 5).

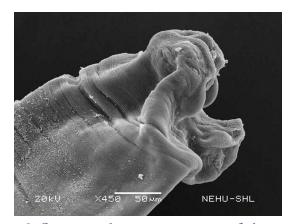


Figure 3: Scanning electron microscopy of the anterior part of *A. galli* treated with PZQ/PTP. Deflation and distortion of the lips are prominent.

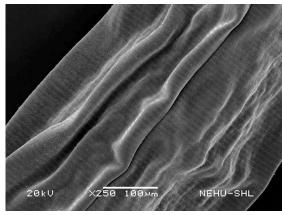


Figure 4: Scanning electron microscopy of the main body of *A. galli* treated with PZQ/PTP. Large longitudinal folds are seen on the cuticle.

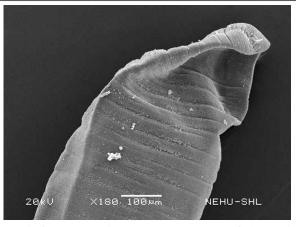


Figure 5: Scanning electron microscopy of the tail end of *A. galli* treated with PZQ/PTP. The tail is deflated. A small terminal spine is seen at the tip.

ABZ-treated nematode revealed finer cuticular shrinkage. As shown in Figure 6, the cephalic region with the lips and surrounding cuticle are all shrunken. The cuticular annulations are also evidently contracted. The cuticular folds are much smaller than those observed on PZQ/PTP-treated nematode (Figure 7). The tail end with an anal opening are also severely shrunken (Figure 8).



Figure 6: Scanning electron microscopy of the anterior part of *A. galli* treated with ABZ. The lips are deflated, shrunken and surrounded by contracted cuticle.

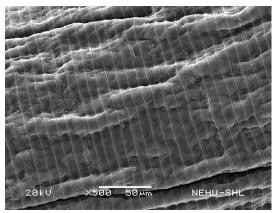


Figure 7: Scanning electron microscopy of the main body of *A. galli* treated with ABZ. The cuticle developed short but many longitudinal folds.

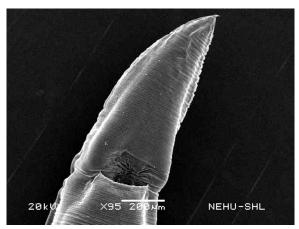


Figure 8: Scanning electron microscopy of the tail end of *A. galli* treated with ABZ. Deflation is very slight but cuticular folds can be seen. The terminal spine is intact.

Table 1: Efficacy of praziquantel/pyrantel pamoate (PZQ/PTP) and albendazole (ABZ) against the poultry nematode, *A. galli*.

Treat- ment	Dose (mg/ml)	Normalised survival time in hr (± SD)	t value against control	t value between drugs
Control	0	100.00 ± 1.21	-	-
PZQ/ PTP	1.25	096.73 ± 1.56	004.05*	049.51*
	2.5	090.44 ± 1.58	011.80*	071.05*
	5	086.98 ± 1.09	019.62*	113.10*
	10	076.01 ± 1.28	033.34*	094.53*
	20	062.28 ± 1.11	056.43*	124.50*
ABZ	1.25	056.27 ± 1.25	061.64*	049.51*
	2.5	037.90 ± 0.89	101.33*	071.05*
	5	025.17 ± 0.78	127.57*	113.10*
	10	007.96 ± 1.21	131.98*	094.53*
	20	001.77 ± 0.44	187.27*	124.50*

^{*}Significantly different at p < 0.05; n: 6.

DISCUSSION

Nematodes are the only group of parasites endowed with thick protective body covering, the cuticle. The nematode cuticle is a syncytium of proteins, collagen being the major component. It acts not only as a protective layer but also as the structural framework of the body (Maizels et al., 1993; Decraemer et al., 2003). Thus, it is in a truest sense a durable and anisometric exoskeleton that maintains the rigid contour of the body while allowing flexibility for movement (Basyoni and Rizk, 2016). It is highly resistant to most chemicals including the digestive enzymes of the host. Although, nematodes are the only helminths with proper mouth and alimentary tract, cuticle still serves as the main entry site for anthelmintic drugs. Drugs are

known to enter by passive diffusion through the cuticle as in other helminths (Mottier et al., 2006; Lanusse et al., 2016; Abongwa et al., 2017).

In nematodes, it is obvious that anthelmintic drugs interfere with the structural integrity of the cuticle by targeting the cuticular proteins as a primary mode of action (Page et al., 2014). Yet different drugs exhibit different efficacy owing to their differential capacities of diffusion, and no drug is 100% effective on nematodes (Stepek et al., 2006). For instance, ABZ is one of the most lipophilic drugs, and thus one of the most diffusible drugs. It has been shown than ABZ has higher efficacy against *Haemonchus contortus* and *Ascaris suum* due to its higher transcuticular diffusion compared to other benzimidazoles (BZDs) or its derivatives (Alvarez et al., 2007).

Our findings also reveal that ABZ directly attacks the cuticle of A. galli. Disintegration of the cuticle is a definitive sign of the anthelmintic effect. In a similar pattern, nitazoxanide acts through the cuticle and was shown to kill the adults and microfilariae of Brugia malayi by destroying the cuticular organisation (Rao et al., 2009). Our study revealed that the extent of damage caused by ABZ on nematode cuticle is quite minimal, implying that the drug acts mostly on the underlying tissues and muscles to cause paralysis and the ensuing death. This supports the fact that ABZ is an established inhibitor of β -tubulin (Alvarez et al., 2007).

We have shown that PTP on the other hand caused extensive cuticular disintegration throughout the body. The cephalic organs are completely shrunk, and the general body is also severely constricted. These indicate that the main target of the drug is the cuticular proteins. However, PTP can also act on the underlying muscle and the intestine as demonstrated in Toxcara canis (Mackenstedt et al., 1993). It is unique in that it can be absorbed both through the cuticle and the alimentary tract via the mouth (Kopp et al., 2008). It is important to note that the cuticular damages seen on A. galli is due to PTP alone, since PZQ is established to be ineffective on nematodes because it is not diffusible through the cuticle. PZQ, with relatively slow uptake through the mouth in the nematodes, enhances the activity of combined drugs by acting synergistically (Southworth et al., 1996; Piantavini et al., 2014). Thus, the vermicidal effects of PTP on A. galli could have been augmented by PZQ by acting on internal tissues other than the cuticle.

CONCLUSION

Albendazole and praziquantel/pyrantel pamoate are commonly used drugs for helminthiases involving infections with cestodes, trematodes, and nematodes. We



show here that both drugs are effective against the intestinal nematode, *A. galli*. For the PZQ/PTP combination drug, the anthelmintic effect on the nematode appeared mostly due to PTP as PZQ is not effective in nematodes. The synergistic effect of PZQ cannot be ruled out, but the drug did not target the cuticle, if at all. ABZ was more effective although it caused less extensive damage on the cuticle of the nematode. This suggests that ABZ mostly act on the sub-cuticular and underlying muscle tissues. Our findings support that the two drugs may be prescribed for helminthiasis due to nematodes in poultry, and that ABZ may be the better drug of choice.

ACKNOWLEDGMENT

The study was funded by the Science and Engineering Research Board, Government on India (EMR/2016/004053). PBL is a Senior Research Fellow under the scheme. SEM was done at the Sophisticated Analytical Instrument Facility (SAIF), North-Eastern Hill University, Shillong, India.

AUTHORS CONTRIBUTION

KLC designed the experiments, analysed the data, and wrote the draft. PBL performed the experiments and finished the manuscript.

CONFLICT OF INTEREST

The authors have declared no conflict of interest.

REFERENCES

- Abongwa M, Martin RJ, Robertson AP (2017). A brief review on the mode of action of antinematodal drugs. Acta Vet. 67: 137–152. https://doi.org/10.1515/acve-2017-0013
- Alvarez LI, Mottier ML, Lanusse CE (2007). Drug transfer into target helminth parasites. Trends Parasitol. 23: 97–104. https://doi.org/10.1016/j.pt.2007.01.003
- Basyoni MM, Rizk EM (2016). Nematodes ultrastructure: complex systems and processes. J. Parasit. Dis. 40: 1130–1140. https://doi.org/10.1007/s12639-015-0707-8
- Bharat GA, Kumar NP, Subhasish B, Ria B (2017). A report of Ascaridia galli in commercial poultry egg from India. J. World's Poult. Res. 7: 23–26.
- •Brar RS, Kumar R, Leishangthem GD, Banga HS, Singh ND, Singh H (2016). *Ascaridia galli* induced ulcerative proventriculitis in a poultry bird. J. Parasit. Dis. 40: 562–564. https://doi.org/10.1007/s12639-014-0509-4
- Decraemer W, Karanastasi E, Brown D, Backeljau T (2003).
 Review of the ultrastructure of the nematode body cuticle and its phylogenetic interpretation. Biol. Rev. 78: 465–510. https://doi.org/10.1017/S1464793102006115
- Foka ES, Yondo J, Katte B, Anangmo CN, Mbida M (2019). Ascaridia galli protect Salmonella typhimurium from antibiotics. South Asian J. Parasitol. 3: 1–5.

- Garcia HH, Lescano AG, Gonzales I, Bustos JA, Pretell EJ, Horton J, Saavedra H, Gonzalez AE, Gilman RH, Tsang VC (2016). Cysticidal efficacy of combined treatment with praziquantel and albendazole for parenchymal brain cysticercosis. Clin. Infect. Dis. 62: 1375–1379. https://doi.
- Gauly M, Homann T, Erhardt G (2005). Age-related differences of *Ascaridia galli* egg output and worm burden in chickens following a single dose infection. Vet. Parasitol. 128: 141–148. https://doi.org/10.1016/j.vetpar.2004.11.023

org/10.1093/cid/ciw134

- •Hong ST (2018). Albendazole and praziquantel: review and safety monitoring in Korea. Infect. Chemother. 50: 1–10. https://doi.org/10.3947/ic.2018.50.1.1
- Kilpinen O, Roepstorff A, Permin A, Nørgaard-Nielsen G, Lawson LG, Simonsen HB (2005). Influence of *Dermanyssus* gallinae and Ascaridia galli infections on behaviour and health of laying hens (Gallus gallus domesticus). Br. Poult. Sci. 46: 26–34. https://doi.org/10.1080/00071660400023839
- Kopp SR, Kotze AC, McCarthy JS, Traub RJ, Coleman GT (2008). Pyrantel in small animal medicine: 30 years on. Vet. J. 178: 177–184. https://doi.org/10.1016/j.tvjl.2007.06.021
- Kyne GM, Curtis MP, Keiser J, Woods DJ (2019). Soil-transmitted helminthiasis-challenges with discovery of novel anthelmintics. In: Swinney D, Pollastri P, Mannhold R, Buschmann H, Holenz J (eds), Neglected Tropical Diseases: Drug Discovery and Development. Wiley-VCH Verlag GmbH and Co. KGaA, Weinheim, Germany, pp. 227–251. https://doi.org/10.1002/9783527808656.ch9
- Lanusse CE, Alvarez LI, Lifschitz AL (2016). Gaining insights into the pharmacology of anthelmintics using *Haemonchus* contortus as a model nematode. Adv. Parasitol. 93: 465–518. https://doi.org/10.1016/bs.apar.2016.02.014
- Mackenstedt U, Schmidt S, Mehlhorn H, Stoye M, Traeder W (1993). Effects of pyrantel pamoate on adult and preadult *Toxocara canis* worms: An electron microscope and autoradiography study. Parasitol. Res. 79: 567–578. https:// doi.org/10.1007/BF00932241
- Maizels RM, Blaxter ML, Selkirk ME (1993). Forms and functions of nematode surfaces. Exp. Parasitol. 77: 380–384. https://doi.org/10.1006/expr.1993.1096
- Moser W, Schindler C, Keiser J (2017). Efficacy of recommended drugs against soil transmitted helminths: systematic review and network meta-analysis. BMJ. 358: j4307. https://doi. org/10.1136/bmj.j4307
- Moser W, Schindler C, Keiser J (2019). Drug combinations against soil-transmitted helminth infections. Adv. Parasitol., 103: 91–115. https://doi.org/10.1016/bs.apar.2018.08.002
- Mottier L, Alvarez L, Ceballos L, Lanusse C (2006). Drug transport mechanisms in helminth parasites: passive diffusion of benzimidazole anthelmintics. Exp. Parasitol. 113: 49–57. https://doi.org/10.1016/j.exppara.2005.12.004
- Page AP, Stepek G, Winter AD, Pertab D (2014). Enzymology of the nematode cuticle: A potential drug target? Int. J. Parasitol. 4: 133–141. https://doi.org/10.1016/j.ijpddr.2014.05.003
- •Piantavini MS, Pontes FL, Uber CP, Stremel DP, Sena MM, Pontarolo R (2014). Chemometric quality inspection control of pyrantel pamoate, febantel and praziquantel in veterinary tablets by mid infrared spectroscopy. Spectrochim. Acta A Mol. Biomol. Spectrosc. 125: 396–403. https://doi.org/10.1016/j.saa.2014.01.078
- Rao RU, Huang Y, Fischer K, Fischer PU, Weil GJ (2009). Brugia malayi: Effects of nitazoxanide and tizoxanide



- on adult worms and microfilariae of filarial nematodes. Exp. Parasitol. 121: 38–45. https://doi.org/10.1016/j.exppara.2008.09.020
- Ruhnke I, Andronicos NM, Swick RA, Hine B, Sharma N, Kheravii SK, Wu SB, Hunt P (2017). Immune responses following experimental infection with *Ascaridia galli* and necrotic enteritis in broiler chickens. Avian Pathol. 46: 602– 609. https://doi.org/10.1080/03079457.2017.1330536
- Schulz JD, Moser W, Hürlimann E, Keiser J (2018).
 Preventive chemotherapy in the fight against soil-transmitted helminthiasis: achievements and limitations.
 Trends Parasitol. 34: 590–602. https://doi.org/10.1016/j.pt.2018.04.008
- Siles-Lucas M, Casulli A, Cirilli R, Carmena D (2018).
 Progress in the pharmacological treatment of human cystic and alveolar echinococcosis: Compounds and therapeutic targets. PLoS Neglect. Trop. Dis. 12: e0006422. https://doi.org/10.1371/journal.pntd.0006422

- Southworth J, Harvey C, Larson S (1996). Use of praziquantel for the control of *Moniezia expansa* in lambs. N. Z. Vet. J. 44: 112–115. https://doi.org/10.1080/00480169.1996.35947
- Stepek G, Buttle DJ, Duce IR, Behnke JM (2006). Human gastrointestinal nematode infections: are new control methods required? Int. J. Exp. Pathol. 87: 325–341. https:// doi.org/10.1111/j.1365-2613.2006.00495.x
- Tinkler SH (2019). Preventive chemotherapy and anthelmintic resistance of soil-transmitted helminths—Can we learn nothing from veterinary medicine? One Health. 31: 100106. https://doi.org/10.1016/j.onehlt.2019.100106
- •WHO (2020a). Schistosomiasis. World health organization fact sheet. Geneva, world health organization. https://www.who.int/news-room/fact-sheets/detail/schistosomiasis
- •WHO (2020b). Soil-transmitted helminth infections. World Health Organization Fact Sheet. Geneva, World Health Organization.https://www.who.int/news-room/fact-sheets/detail/soil-transmitted-helminth-infections

